

# Use and Diversity of Medicinal Plants in Aquaculture Practices

Elias Fernandes de Medeiros Júnior<sup>1,\*</sup>, Eugênio Bispo da Silva Júnior<sup>1</sup>, Xenusa Pereira Nunes<sup>1</sup>, Rosimeire Morais Cardeal Simão<sup>1</sup>, Cleoni Virgínio da Silveira<sup>2</sup>, Luciana Souza de Oliveira<sup>2</sup>, Lucia Marisy Souza Ribeiro de Oliveira<sup>3</sup>, Denes Dantas Vieira<sup>3</sup>, Xirley Pereira Nunes<sup>3</sup>

<sup>1</sup>PhD students from the doctoral graduate program in Agroecology and Territorial Development, Federal University of the São Francisco River Valley (UNIVASF)-Brazil

<sup>2</sup> PhD Professor, of the IFAM-Campus São Gabriel da Cachoeira-AM

<sup>2</sup>PhD Professor, Federal Institute of Education, Science and Technology of Sertão Pernambucano, Campus Petrolina Zona Rural, Brazil

<sup>3</sup>PhD Professor, doctoral graduate program in Agroecology and Territorial Development, Federal University of the São Francisco River Valley (UNIVASF)-Brazil

\*Corresponding Author

**Abstract**—Medicinal plants are widely used in Brazilian folk medicine because of their various properties. Their chemical components have attracted the attention of researchers from the field of animal production, particularly aquaculture. This study aimed to analyze the use of medicinal plants in aquaculture practices. The main results showed that medicinal plants have been used in the management of anesthesia, in antibacterial, antifungal, and antiparasitic treatments, and as growth-promoting additives. When the potential of Brazilian flora is considered, little knowledge has been gained about the use of medicinal plants in aquaculture practices, especially when considering the native species, which may represent an important research front in the development of phytopharmaceuticals for the treatment of diseases of cultured aquatic organisms.

**Keywords**—Fish farming, Plant extract, Tambaqui, Tilapia.

## I. INTRODUCTION

In 2016, the world aquaculture production, including aquatic plants, was 110.2 million tons (FAO, 2018). In this context of growing aquaculture activities, Brazilian fish farming reported an 8% growth in 2017, with a total production of 691,700 tons of farmed fish (Peixe & Peixe, 2018). Success in fish farming depends on a number of management practices, including water quality control, quarantine of newly acquired batches, and providing quality and balanced food. This ensures fish health and, consequently, disease prevention (Santos et al., 2013).

According to Tavechio et al. (2009), many of the diseases affecting fish farming are caused by infectious agents that can make the activity costly and unprofitable due to high mortality during outbreaks of infection/infestation. Natural products have been the objects of growing attention, representing a potential alternative as antibacterial agents in the culture of aquatic

organisms, mainly because of their easy availability, relatively low cost, and proven efficiency against a number of pathogens (Galina, Yin, Ardó, & Jeney, 2009; Nazzaro, Fratianni, De Martino, Coppola, & De Feo, 2013).

Herbal medicines and compounds from plants (phytopharmaceuticals) can be applied as prophylactic or therapeutic measures against bacterial diseases in aquatic organisms, mainly using two methods: through bathing or by incorporating in the feed (Saccol et al., 2013; Sutili et al., 2014). The substances found in essential oils and plant extracts can act directly on bacteria, causing cell lysis, inhibiting the antibacterial activity of other substances, or inhibiting bacterial resistance mechanisms and virulence factors (Stavri, Piddock, & Gibbons, 2007).

Thus, considering the nutritional, immunological, bactericidal, antifungal, anthelmintic, and antiparasitic properties of medicinal plants, among other benefits, this

study aimed to analyze the use of medicinal plants in aquaculture practices.

## II. MATERIALS AND METHODS

This study was conducted from August to September 2019; it is a qualitative and quantitative review of the literature about the use of medicinal plants in aquaculture practices. Data were searched in three databases: Google Scholar, SciELO, and the journal database of the Brazilian Coordination for the Improvement of Higher-Education Personnel (CAPES), using “AND” as the Boolean operator. The keywords or indexing terms were “medicinal plants AND aquaculture,” “medicinal plants AND fish farming,” “medicinal plants AND tambaqui (*Colossoma macropomum*),” and “medicinal plants AND tilapia (*Oreochromis niloticus*).” The latter two searches included the scientific name of the species most commonly produced in aquaculture freshwater in Brazil.

The inclusion criteria were scientific articles published in scientific journals in the last 10 years only (2009–2019). Duplicate articles or articles not related to the study theme, as well as abstracts in annals of scientific journals, dissertations, theses, technical newsletters, and other documents were excluded. The data collected from the articles were inserted into Microsoft Office Excel® 2010 spreadsheets for content analysis and data tabulation.

## III. RESULTS AND DISCUSSION

The search in the SCIELO database found only one article based on the descriptor “medicinal plants AND

aquaculture,” one article for “medicinal plants AND fish farming,” no results for “medicinal plants AND tambaqui (*Colossoma macropomum*),” and two results for “medicinal plants AND tilapia (*Oreochromis niloticus*).” In CAPES journals, six articles were found for “medicinal plants AND aquaculture,” but only one met the inclusion criteria; ten results were found for “medicinal plants AND fish farming,” but only two were related to the study theme; and two articles each were found for both “medicinal plants AND tambaqui” and “medicinal plants AND tilapia.”

In the Google Scholar database, 2,300 articles were found for “medicinal plants AND aquaculture,” 2,850 for “medicinal plants AND fish farming,” 549 for “medicinal plants AND tambaqui (*Colossoma macropomum*),” and 1,360 for “medicinal plants AND tilapia (*Oreochromis niloticus*).” After a careful analysis of all Google Scholar search results, only 28 items were related to the study theme, and they were selected for content analysis.

The medicinal plants most commonly used in aquaculture practices were pitanga (*Eugenia uniflora* L.), *Hyptidendron canum* (Pohl ex Benth.) Harley, clove basil (*Ocimum gratissimum* L.), mint (*Mentha arvensis* L.), clove (*Eugenia caryophyllus* L.), passion flower (*Passiflora incarnata* L.), aloe vera (*Aloe vera* (L.) Burm. f.), lemon balm (*Lippiaspp.* L.) (Verbenaceae), cockspur coral tree (*Erythrina crista-galli* L.), velame (*Croton heliotropiifolius* Kunth), country almond (*Terminalia catappa* L.), yellow cinnamon (*Nectandra grandiflora* Ness), and plants from the Piperaceae family. Table 1 shows the use of these medicinal plants in aquaculture practices.

Table 1. List of medicinal plants used in aquaculture and their applications

Medicinal plants	Effect/use in aquaculture	Fish species	Reference
Pitanga ( <i>Eugenia uniflora</i> L.)	Clinical study	<i>Oreochromis niloticus</i>	Fiuza et al. (2011)
<i>Hyptidendron canum</i> (Pohl ex Benth.) Harley	Clinical study	<i>Oreochromis niloticus</i>	Fiuza et al. (2015)
Clove basil ( <i>Ocimum gratissimum</i> L.)	Anesthesia	<i>Brycon amazonicus</i>	Ribeiro et al. (2016)
	Anesthesia	<i>Colossoma macropomum</i>	Façanha & Gomes (2005)
Mint ( <i>Mentha arvensis</i> L.)	Anesthesia	<i>Xiphophorus maculatus</i>	Hoshiba et al. (2015)
	Anesthesia	<i>Centropomus parallelus</i>	Souza et al. (2012)
	Anesthesia	<i>Piaractus mesopotamicus</i>	Gonçalves et al. (2008)
Clove ( <i>Eugenia caryophyllus</i> L.)	Anesthesia	<i>Piaractus mesopotamicus</i>	Gonçalves et al. (2008)
	Anesthesia	<i>Centropomus parallelus</i>	Souza et al. (2012)

Passion flower ( <i>Passiflora incarnata</i> L.)	Nutrition	<i>Oreochromis niloticus</i>	Oliveira, Pereira-Da-Silva, & Bueno (2010)
Aloe vera ( <i>Aloe vera</i> (L.) Burm. f.)	Semen cryopreservation	<i>Colossoma macropomum</i>	Melo-Maciel et al. (2015)
Lemon balm ( <i>Lippia</i> spp.L.)	Nutrition	<i>Oreochromis niloticus</i>	Rodrigues-Soares et al. (2018)
	Antibacterial	<i>Colossoma macropomum</i>	Oliveira et al. (2018)
Cockspur coral tree ( <i>Erythrina crista-galli</i> L.)	Anesthesia	<i>Carassius auratus</i>	Siqueira, Sousa, Tirloni, & Gebara(2019)
<i>Velame</i> ( <i>Croton heliotropifolius</i> Kunth)	Nutrition	<i>Oreochromis niloticus</i>	Souza et al. (2018)
	Nutrition	<i>Oreochromis niloticus</i>	Santos et al. (2015)
Country almond ( <i>Terminalia catappa</i> L.)	Antiparasitic	<i>Colossoma macropomum</i>	Claudiano et al. (2009)
	Behavior/Performance	<i>Betta splendens</i>	Santos et al. (2013)
Yellow cinnamon ( <i>Nectandra grandiflora</i> Ness)	Antiparasitic	—	Rodrigues et al. (2017)
Piperaceae (spp.)	Anthelmintic	<i>Colossoma macropomum</i>	Santos et al. (2018)

Source: The authors themselves (2019).

### 3.1 Pitanga (*Eugenia uniflora* L.)

The *pitanga* tree (*E. uniflora* L.) has edible fruits that are well known and appreciated in Brazil; the infusion of its leaves has applications in folk medicine mainly as a hypotensive, anti-gout, stomachic, and hypoglycemic agent (Auricchio & Bacchi, 2003). In vitro and in vivo tests have shown that extracts from the leaves of *E. uniflora* L. have several pharmacological properties, such as antidiarrheal, diuretic, anti-inflammatory, and antifungal actions (Almeida, Faria, & Silva, 2012). Fiuza et al. (2011) found that tilapia (*Oreochromis niloticus*) exposed to different *E. uniflora* L. extracts and leaf fractions had a vasodilation effect on gills, and toxic effects, such as detachment and desquamation of the respiratory epithelium and hyperplasia of the interlamellar epithelium cells. These effects were more pronounced in those individuals who received the highest concentrations. This study contributed to establish *Nile tilapia* as a model system for testing active ingredients of plants, such as the vasodilation effect, the effect on cell morphology and on the tissues of the gills, the main respiratory organ of fishes.

### 3.2 *Hyptidendron canum* (Pohl ex Benth) Harley

*Hyptidendron* belongs to the Lamiaceae family, which consists of herbs, shrubs, and trees, with usually quadrangular branches (Fiuza et al., 2010). *Hyptidendron canum* is native to the Brazilian savannah (*cerrado*) regions and is frequently used in alternative medicine for

the treatment of malaria, with anti-inflammatory, anti-ulcerative, and anti-hepatotoxic actions (Brandão, 1991; Ferri & Ferreira, 1992). Desquamation of the respiratory epithelium, changes in curvature, cell hyperplasia, and vasodilation in the lamellae and in the central vessel of the filaments were the main results that Fiuza et al. (2015) found in the gills of *O. niloticus* submitted to ethanol extract and to hexane, chloroform, and ethyl acetate fractions of *H. canum*. The authors pointed out that *H. canum* caused inflammatory processes and injuries that varied according to the dose administered.

### 3.3 Clove basil (*Ocimum gratissimum* L.)

*Ocimum gratissimum*, popularly known as clove basil, belongs to the family Lamiaceae, one of the largest families of angiosperms (Cruz & Bezerra, 2017). In traditional medicine, *O. gratissimum* has several indications, such as an anti-flu bath, and diuretic, febrifugal, anti-bleeding, antifungal, antioxidant, antibacterial, antidiarrheal, hypoglycemic, and anti-inflammatory indications (Lemos et al. 2005; Stanley, Ifeanyi, Chinedum, & Chinenye, 2014). The use of *O. gratissimum* in aquaculture activities is associated with the presence of eugenol, the main active component present in the plant, which has been studied as a natural anesthetic used in fish farming. Ribeiro et al. (2016) used the essential oil of clove basil as a natural anesthetic in young *matrinxã* (*Brycon amazonicus*), a neotropical fish found in the Amazon

region, without side effects on the fish; as no death was reported for 30 days, the authors suggested that the essential oil of *O. gratissimum* did not pose any risk for handlers at a concentration of 20–60 mg.L<sup>-1</sup> for a 10-min exposure, thus allowing its use in *matrinxã* anesthesia.

### 3.4 Mint (*Mentha arvensis* L.)

The *Mentha* genus comprises approximately 25 different species of mints and related plants that belong to the Lamiaceae family (Watanabe, Nosse, Garcia, & Pinheiro Povh, 2006). Mint (*Mentha arvensis* L.) is an aromatic plant, with menthol as the substance found in the largest amounts in the composition of its essential oil (Paulus et al. 2007; Arrigoni-Blank, 2011). Menthol is a natural anesthetic, which has been used for different species grown in Brazil, proving to be efficient and safe in fish anesthesia procedures, mainly in the following species: tambaqui (*Colossoma macropomum*) (Façanha & Gomes, 2005); platys (*Xiphophorus maculatus*) (Hoshiba et al. 2015); *robalo-peva* (*Centropomus parallelus*), and *pacu* (*Piaractus mesopotamicus*) (Gonçalves, Santos, Fernandes, & Takahashi, 2008).

### 3.5 Clove (*Eugenia caryophyllus* L.)

The clove plant belongs to the Myrtaceae family, which has approximately 3,000 species of tropical and subtropical trees and shrubs. Its scientific name varies with its classification; it was recently classified as *Syzygium aromaticum* (L.) Merr. et Perry, despite several previous citations, as follows: *Eugenia caryophyllus* (Sprengel) Bullock et Harrison, *Caryophyllus aromaticus* L., *E. caryophyllata* Tumb, and *E. aromatica* (L.) Baill (Maeda, Bovi, Bovi, & Lago, 1990). Eugenol, the main component of the plant, has anti-inflammatory, healing, and analgesic effects, and it is effective in reducing bacteria present in the mouth (Silvestri et al., 2010). The use of cloves in aquaculture is related to the need to anesthetize cultivated organisms for performing management practices, such as biometrics, reproduction, and transport. The use of cloves has been recommended for *robalo-peva* (Souza et al., 2012) and *pacu* (Gonçalves et al., 2008) juveniles, proving to be safe and efficient for the animals handled.

### 3.6 Passion flower (*Passiflora incarnata* L.)

Passion flower, of the *Passiflora incarnata* species, has the potential to reduce stress. Its activity is related to the presence of pyronic derivatives, harman alkaloids, and flavonoids, to which sedative and anxiolytic effects are attributed (Dhawan, Kumar, & Sharma, 2003). In a study conducted by Oliveira et al. (2010), the authors suggested that passion flower extract could be included in the diet of young tilapia, without prejudice to food consumption and growth. They also reported that the extract changed the

morphometry of hepatocytes, suggesting the activity of flavonoids on carbohydrate metabolism, which contributed to increased glycogen levels in liver, particularly in the group that received 100 mg.kg<sup>-1</sup> of the extract.

### 3.7 Aloe vera (*Aloe vera* (L.) Burm. f.)

*Aloe vera* belongs to the Aloaceae family, which includes approximately 15 genera and 800 species. It is a herbaceous plant that grows on any type of soil, but it is better adapted to light and sandy soils and does not require much water (Freitas, Rodrigues, & Gaspi, 2014). It is very common in Brazil, where it is popularly used in wound healing, in the treatment of burns, conjunctivitis, rheumatic pain, and other uses (Guerra, Araújo, & Oliveira, 2008; Araújo, Lemos, Menezes, Fernandes, & Kenrtopf, 2015). The use of *Aloe vera* in fish farming was studied by Melo-Maciel et al. (2015) in tambaqui semen cryopreservation, and the authors concluded that *A. vera* as a crude extract did not improve sperm production during the cryopreservation process.

### 3.8 Lemon balm (*Lippia* spp. Linn.)

The *Lippia* genus (Verbenaceae) includes approximately 200 species of herbs, shrubs, and small trees mainly found in Central America and in the tropical regions of Africa, North America, South America, and Australia (Reis et al. 2014; Gomes, Nogueira, & Moraes, 2011). The species of this genus are widely used in folk medicine because they have anti-inflammatory, antifungal, antiseptic, antihypertensive, anxiolytic, anti-*Leishmania*, antiviral, and digestive properties, among other applications (Soares & Tavares-Dias, 2013; Costa, Souza, Brito, & Fontenelle, 2017). Rodrigues-Soares et al. (2018) added *Lippia alba* essential oil to the feed of tilapia (*O. niloticus*) in order to analyze the hemato-immunological parameters. The authors concluded that the essential oil did not contribute to anti-inflammatory activities; however, an increase in the number of neutrophils was observed. In a study by Oliveira et al. (2018), the authors used *L. organoides* essential oil to control infections by *Aeromonas hydrophila* in tambaqui (*C. macropomum*) juveniles and found a survival rate of 79.2% after a therapeutic bath with 10 mg.L<sup>-1</sup> of essential oil; the changes in hematological and biochemical parameters were not significant.

### 3.9 Cockspur coral tree (*Erythrina crista-galli* L.)

The *Erythrina* genus (Fabaceae) contains more than 100 species distributed in the tropical and subtropical areas of the Americas, Africa and Australia (Kone, Solange, & Dosso, 2011). This tree is found in very humid areas, in secondary open formations, from Maranhão to Rio Grande do Sul (Gratieri-Sossella & Nienow, 2008).

Pharmacological investigations have demonstrated that *E.crista-galli* seed extracts have sedative, hypertensive, laxative, and diuretic properties (Maier, Rödi, Deus-Neumann, & Zenk,1999). Additionally, its bark is used to treat many diseases associated with rheumatism and hepatitis (Hashimoto, 1996). Siqueira et al. (2019) conducted a study to evaluate the anxiolytic effect of *E. crista-galli* extract in goldfish (*Carassius auratus*) juveniles, and the authors observed that the extract did not present fish toxicity or mortality. However, it did cause undesirable changes in blood physiological parameters when used in doses above 100 mg.L<sup>-1</sup>. Thus, the authors reported that the anxiolytic action was not beneficial to the fish species analyzed.

### 3.10 *Velame* (*Croton heliotropifolius* Kunth)

*Croton* is the second largest genus of Euphorbiaceae, with approximately 1,200 species predominantly distributed across the American continent (Berry, Hipp, Wurdack, Van, & Riina, 2005). Brazil has the largest number of species, approximately 350, distributed across the most diverse ecosystems, especially the savannah (*cerrado*), the semiarid plateau (*caatinga*), and rupestrian fields (Berry et al. 2005). This genus' species have demonstrated anti-inflammatory (Ramos et al., 2013), gastroprotective (Coelho-De-Souza et al., 2013), and woundhealing (Cavalcanti et al., 2012) properties. Souza et al. (2018) evaluated the nutritional effect of adding extracts of *Croton heliotropifolius* to the diet of young tilapia. The authors found that the inclusion of *velame* extract reduced the average weight gain and the total blood sugar and protein concentrations, whereas the specific growth and survival rates had a small improvement with the addition of 2.0% of crude *velame* extract to the diet.

### 3.11 Country almond (*Terminalia catappa* L.)

The country almond (*Terminalia catappa*) belongs to the Combretaceae family. It is an ornamental plant found in several tropical countries, widely used in urban forestry, and present in coastal areas to provide shade (Francis, 1989). Its origin is the coastal areas of eastern India, Indochina, Malaysia, northern Australia, Oceania, Philippines, and Taiwan (Francis, 1989). Studies have demonstrated its bactericidal and fungicidal effects (Costa, Bevilaqua, Morais, & Vieira, 2008), and its anti-*Helicobacter pylori* and anti-ulcer (Pinheiro Silva et al., 2015), anti-diabetic (Nagappa, Thakurdesai, Venkat Rao, & Singh, 2003), and anti-inflammatory (Fan et al., 2004) properties. *T. catappa* leaves were tested as a growth-promoting additive for *N. tilapia* (*Oreochromis niloticus*) juveniles, and the results did not show any positive effects on performance. However, when

administered at high levels, it reduced mortality (Santos et al., 2015). In a study conducted by Claudiano et al. (2009), an aqueous extract of dry leaves of *T. catappa* presented improved efficiency at the dose of 120 ml.L<sup>-1</sup> and effectively controlled monogenetic parasites and the protozoan *Piscinoodinium pillulare* in *tambaqui* (*C. macropomum*) juveniles; however, it had no effect against the protozoan *Ichthyophthirius multifiliis*. Santos et al. (2013) evaluated the performance and behavior of *Betta splendens* fish growing at different concentrations of *T. catappa* aqueous extract, and concluded that the extract had no effect on fish performance, whereas fish behavior was influenced by the concentration—that is, higher concentrations caused the fish to be calmer.

### 3.12 Yellow cinnamon (*Nectandra grandiflora* Ness)

The yellow cinnamon (*Nectandra grandiflora* Ness) (Lauraceae) is an endemic species of Brazil, predominantly found in the Atlantic rainforest and *cerrado* biomes (Lorenzi & Brasileira, 2002). Regarding the properties of this medicinal plant, Ribeiro (2002) and Ribeiro et al. (2005) analyzed the antioxidant activity of an ethanol extract of *N. grandiflora* leaves. The extract inhibited the oxidation of  $\beta$ -carotene, analgesic (da Silva-Filho et al., 2004), antibacterial (Ferraz et al., 2018), and anxiolytic (Garlet et al., 2019) actions. Rodrigues et al. (2017) reported an in vitro analysis in which the essential oil of *N. grandiflora* exhibited an antiparasitic effect against *I. multifiliis*, a fish parasite that causes considerable losses in aquaculture. They also suggested that in vivo studies should be conducted to develop a product for the control of *I. multifiliis*.

### 3.13 Species of the Piperaceae family

The *Piper* genus belongs to the Piperaceae family, which was described in the 18<sup>th</sup> century by Linnaeus. It has approximately 12 genera, of which *Piper* and *Peperomia* are the most important ones in the Brazilian flora (Medeiros & Guimarães, 2007). This family's pharmacological potential is related to its antitumor (Duh, Wu, & Wuang, 1990), insecticide (Mamood, Hidayatulfathi, Budin, Ahmad Rohi, & Zulfakar, 2017), anesthetic (López et al., 2016), anesthetic, and antimycobacterial (Cunico et al., 2015) properties. Santos et al. (2018) found that essential oils from four species of plants of the family Piperaceae—*Piper hispidinervum*, *P. hispidum*, *P. marginatum*, and *P. callosum*—showed anthelmintic efficacy against the acanthocephalon parasite *Neoechinorhynchus buttnerae* in *tambaqui* (*C. macropomum*) juveniles, and that parasite mortality occurred at the highest concentrations and times of exposure to the essential oils. The authors concluded the

essential oils were an alternative source for direct use or for the development of anthelmintic herbal medicines, requiring advanced studies for in vivo treatment.

#### IV. CONCLUSION

This study on the use of medicinal plants in aquaculture practices demonstrated the diversity of plants used. However, when analyzing the biodiversity of the Brazilian flora across its most diverse biomes, the production of knowledge in this area still presents few results, particularly considering that the tests were conducted with *Nile tilapia*, the main exotic species grown in Brazilian aquaculture.

The potential of the Brazilian flora for the production of phytopharmaceuticals that can be used in aquaculture still needs further investigation, mainly regarding its use with cultivated native species. While fish farming activities have increased in Brazil, many cases of diseases related to the presence of parasites, endoparasites, bacteria, fungi, and viruses have been reported because of the intensive systems used in fish farming practices. Fish sanitary diagnoses are usually expensive, particularly for small fish farmers. Thus, the use of medicinal plants, especially those producing essential oils and mentioned in this study, has presented great potential for the treatment of aquaculture diseases. Further studies evaluating their use are required to reduce the costs involved in the treatment of these diseases.

#### ACKNOWLEDGMENTS

We/The authors thank Crimson Interactive Pvt. Ltd. (Ulatu) – [www.ulatus.com.br](http://www.ulatus.com.br) for their assistance in manuscript translation and editing.

#### REFERENCES

- [1] Organización de las Naciones, F. A. O. *Unidas para la Alimentación y la Agricultura. El Estado mundial de la pesca y la acuicultura. Producción de la acuicultura*. (2018).
- [2] Peixe, B. R., & Peixe, A. (2018). Associação Brasileira da Piscicultura. BR da piscicultura, 2018.
- [3] Santos, D. M., Santos, E. L., Souza, A. P. L., Temoteo, M. C., Cavalcanti, M. C. A., Silva, F. C. B., & Pontes, E. C. (2013). Uso do extrato aquoso da folha desidratada de amendoeira (*Terminalia catappa*) no cultivo de *Bettasplendens*. PUBVET, Londrina. *Art*, 7(4), Ed. 227, 1505.
- [4] Tavechio, W. L. G., Guidelli, G., & Portz, L. (2009). Alternativas para a prevenção e o controle de patógenos em piscicultura. *Boletim do Instituto de Pesca, São Paulo*, 35(2), 335–341.
- [5] Galina, J., Yin, G., Ardó, L., & Jeney, Z. (2009). The use of immunostimulating herbs in fish. An overview of research. *Fish Physiology and Biochemistry*, 35(4), 669–676. doi:[10.1007/s10695-009-9304-z](https://doi.org/10.1007/s10695-009-9304-z)
- [6] Nazzaro, F., Fratianni, F., De Martino, L., Coppola, R., & De Feo, V. (2013). Effect of essential oil on pathogenic bacteria. *Pharmaceuticals*, 6(12), 1451–1474. doi:[10.3390/ph6121451](https://doi.org/10.3390/ph6121451)
- [7] Saccol, E. M. H., Uczay, J., Pês, T. S., Finamor, I. A., Ourique, G. M., Riffel, A. P. K., ... Pavanato, M. A. (2013). Addition of *Lippia alba* (Mill.) N. E. Brow essential oil to the diet of the silver catfish: An analysis of growth, metabolic and blood parameters and the antioxidant response. *Aquaculture*, 416–417, 244–254. doi:[10.1016/j.aquaculture.2013.09.036](https://doi.org/10.1016/j.aquaculture.2013.09.036)
- [8] Sutili, F. J., Kreutz, L. C., Noro, M., Gressler, L. T., Heinzmann, B. M., de Vargas, A. C., & Baldisserotto, B. (2014). The use of eugenol against *Aeromonas hydrophila* and its effect on hematological and immunological parameters in silver catfish (*Rhamdia quelen*). *Veterinary Immunology and Immunopathology*, 157(3–4), 142–148. doi:[10.1016/j.vetimm.2013.11.009](https://doi.org/10.1016/j.vetimm.2013.11.009)
- [9] Stavri, M., Piddock, L. J., & Gibbons, S. (2007). Bacterial efflux pump inhibitors from natural sources. *Journal of Antimicrobial Chemotherapy*, 59(6), 1247–1260. doi:[10.1093/jac/dkl460](https://doi.org/10.1093/jac/dkl460)
- [10] Auricchio, M. T., & Bacchi, E. M. (2003). Folhas de *Eugenia uniflora* L. (Pitanga): Propriedades farmacobotânicas, químicas e farmacológicas. *Revista do Instituto Adolfo Lutz*, 62(1), 55–61.
- [11] Almeida, D. J., Faria, M. V., & Silva, P. R. (2012). Biologia Experimental em Pitangueira: Uma revisão de cinco décadas de publicações científicas. *Revista do Setor de Ciências Agrárias e Ambientais. Guarapuava*, 8(1), 177–193.
- [12] Fiuza, T. S., Silva, P. C., Paula, J. R., Tresvenzol, L. M. F., Souto, M. E. D., & Sabóia-Morais, S. M. T. (2011). Análise tecidual e celular das brânquias de *Oreochromis niloticus* L. tratadas com extrato etanólico bruto e frações das folhas da pitanga (*Eugenia uniflora* L.)-Myrtaceae. *Revista Brasileira de Plantas Medicinais*, 13(4), 389–395. doi:[10.1590/S1516-05722011000400003](https://doi.org/10.1590/S1516-05722011000400003)
- [13] Fiuza, T. S., Rezende, M. H., Sabóia-Morais, S. M. T., Tresvenzol, L. M. F., Ferreira, H. D., & Paula, J. R. (2010). Estudo das folhas e caule de *Hyptidendron canum* (Pohlex Benth.) Harley, Lamiaceae. *Rev. Bras. Farmacogn*, 20(2, abril–Mai), 192–200. doi:[10.1590/S0102-695X2010000200010](https://doi.org/10.1590/S0102-695X2010000200010)
- [14] Brandão, M. (1991). Plantas medicinais do Cerrado mineiro. *Informativo Agropecuário*, 15, 15–20.
- [15] Ferri, P. H., & Ferreira, H. D. *Fitoquímica das folhas de Hyptis Benth. Semana de Química, Goiânia, GO*, p. 1–32. (1992).
- [16] Fiuza, T. S., Silva, P. C., Paula, J. R., Tresvenzol, L. M. F., Ferreira, H. D., & Sabóia-Morais, S. M. T. (2015). Efeito do extrato etanólico bruto e das frações da *Hyptidendron canum* (Pohlex Benth.) Harley em brânquias de

- Oreochromis niloticus* L. Rev. Bras. PI. Medico, Campinas, 17(1), 1–8.
- [17] Cruz, M. J. F., & Bezerra, S. B. (2017). Obtenção do óleo essencial de *Ocimum gratissimum* L. para o desenvolvimento de cosméticos de limpeza facial. *Revistã Diálogos Acadêmicos, Fortaleza*, 6(2, julho/Dez.).
- [18] Lemos, JdA., Passos, X. S., Fernandes, OdF. L., Paula, J. Rd, Ferri, P. H., Souza, L. K. He, ... Silva, MdR. R. (February 2005). Antifungal activity from *Ocimum gratissimum* L. towards *Cryptococcus neoformans*. *Memórias Do Instituto Oswaldo Cruz*, 100(1), 55–58. doi:[10.1590/S0074-02762005000100011](https://doi.org/10.1590/S0074-02762005000100011)
- [19] Stanley, M. C., Ifeanyi, O. E., Chinedum, O. K., & Chinenye, N. D. (2014). The antibacterial activity of leaf extracts of *Ocimum gratissimum* and *Sida acuta*. *International Journal of Microbiological Research*, v. 5, n. 2, 124–129
- [20] Ribeiro, A. S., Batista, E. D. S., Dairiki, J. K., Chaves, F. C. M., & Inoue, L. A. K. A. (Styczeń–March 2016). Anesthetic properties of *Ocimum gratissimum* essential oil for juvenile matrinxã. *Acta Scientiarum. Animal Sciences*, 38(1), 1–7. doi:[10.4025/actascianimsci.v38i1.28787](https://doi.org/10.4025/actascianimsci.v38i1.28787)
- [21] Watanabe, C. H., Nosse, T. M., Garcia, C. A., & Pinheiro Povh, N. (2006). Extração do óleo essencial de menta (*Mentha arvensis* L.) por destilação por arraste a vapor e extração com etanol. *Rev. Bras. PI med Botucatu*, 8(4), 76–86
- [22] Paulus, D., Medeiros, S. L. P., Santos, O. S., Manfron, P. A., Paulus, E., & Fabbrin, E. (2007). Teor e qualidade do óleo essencial de menta (*Mentha arvensis* L.) produzida sob cultivo hidropônico e em solo. *Rev. Bras. PI med Botucatu*, 9(2), 80–87.
- [23] Arrigoni-Blank, MdF., Costa, A. S., Fonseca, V. O., Alves, P. B., & Blank, A. F. (Styczeń–March 2011). Micropropagação, aclimação, teor e composição química do óleo essencial de genótipos de hortelã japonesa. *Revista Ciência Agronômica*, 42(1), 175–184. doi:[10.1590/S1806-66902011000100022](https://doi.org/10.1590/S1806-66902011000100022)
- [24] Façanha, M. F., & Gomes, LdC. (2005). A eficácia do mentol como anestésico para tabaqui (*Colossomamacropomum*, Characiformes: Characidae). *Acta Amazonica*, 35(1), 71–75. doi:[10.1590/S0044-59672005000100011](https://doi.org/10.1590/S0044-59672005000100011)
- [25] Hoshihira, M. A., Dias, R. M. S., Moreira, K. M. F., Cunha, L., Geraldo, A. M. R., & Tamajusuku, A. S. K. (2015). Cloveoil and menthol as anesthetic for platy. *Boletim do Instituto de Pesca, São Paulo*, 41 (esp), 737–742.
- [26] Gonçalves, A. F. N., Santos, E. C. C., Fernandes, J. B. K., & Takahashi, L. S. (2008). Mentol e eugenol como substituto da benzocaína na indução anestésica de juvenis de pacu. *Acta Scientiarum – Animal Sciences, Maringá*, 30(3), 339–344.
- [27] Maeda, J. A., Bovi, M. L. A., Bovi, O. A., & Lago, A. Ad. (1990). Craveiro-da-Índia: Características físicas das sementes e seus efeitos na germinação e desenvolvimento vegetativo. *Bragantia*, 49(1), 23–36. doi:[10.1590/S0006-87051990000100003](https://doi.org/10.1590/S0006-87051990000100003)
- [28] Silvestri, J. D. F., Paroul, N., Czyewski, E., Lerin, L., Rotava, I., Cansian, R. L., ... Treichel, H. (2010). Perfil da composição química e atividade antibacteriana e antioxidante do óleo essencial do cravo-da-índia (*Eugenia caryophyllata* Thunb.). *Revista Ceres, Viçosa*, 57(5), 589–594, set/out. doi:[10.1590/S0034-737X2010000500004](https://doi.org/10.1590/S0034-737X2010000500004)
- [29] Souza, R. A. R., Carvalho, C. V. A., Nunes, F. F., Scopel, B. R., Guarizi, J. D., & Tsuzuki, M. Y. (2012). Efeito comparativo da benzocaína, mentol e eugenol como anestésico para juvenis de robalo peva. *Boletim do Instituto de Pesca, São Paulo*, 38(3), 247–255.
- [30] Dhawan, K., Kumar, S., & Sharma, A. (2003). Evaluation of central nervous system effects of *Passiflora incarnata* in experimental animals. *Pharmaceutical Biology*, 41(2), 87–91. doi:[10.1076/phbi.41.2.87.14241](https://doi.org/10.1076/phbi.41.2.87.14241)
- [31] Oliveira, R. H. F., Pereira-Da-Silva, E. M., & Bueno, R. S. (2010). BARONE, A.A.C.O extrato de maracujá sobre a morfometria de hepatócitos da tilápia do Nilo. *Ciência Rural*, 40(12, Dez.).
- [32] Freitas, V. S., Rodrigues, R. A. F., & Gaspi, F. O. G. (2014). Propriedades farmacológicas de *Aloe vera* (L.) Burm. f. *Revista Brasileira de Plantas Mediciniais*, 16(2), 299–307. doi:[10.1590/S1516-05722014000200020](https://doi.org/10.1590/S1516-05722014000200020)
- [33] Guerra, M. F. L., Araújo, E. C., & Oliveira, R. A. G. (2008). Uso empírico in natura de aloesp em portadores de conjuntivite. *Revistã de Enfermagem da UFPE*, 2(1), 36–46
- [34] Araújo, M. A., Lemos, I. C. S., Menezes, I. R. A., Fernandes, G. P., & Kenrtopf, M. R. (junio 2015). Uso de plantas medicinais para tratamento de feridas. *R. Interd*, v, 8(2), abr. Mai, 60–67.
- [35] Melo-Maciel, M. A. P., Leite-Castro, L. V., Leite, J. S., Oliveira, M. S., Almeida-Monteiro, P. S., Nunes, J. F., & Salmito-Vanderley, C. S. B. (2015). Aloe vera na criopreservação do sêmen de tambaqui (*Colossomamacropomum*). *Arquivo Brasileiro de Medicina Veterinária e Zootecnia*, 67(3), 945–949. doi:[10.1590/1678-4162-7807](https://doi.org/10.1590/1678-4162-7807)
- [36] Reis, A. C., Sousa, S. M., Vale, A. A., Pierre, P. M. O., Franco, A. L., Campos, J. M. S., ... Viccini, L. F. (2014). *Lippia alba* (Verbenaceae): A new tropical autopolyploid complex? *American Journal of Botany*, 101(6), 1002–1012. doi:[10.3732/ajb.1400149](https://doi.org/10.3732/ajb.1400149)
- [37] Gomes, S. V. F., Nogueira, P. C. L., & Moraes, V. R. S. (2011). Aspectos químicos e biológicos do gênero *Lippia* enfatizando *Lippia gracilis* Schauer. *Eclética Química*, 64–77.
- [38] Soares, B. V., Tavares-Dias, M. (2013). Espécies de *Lippia* (Verbenaceae), seu potencial bioativo e importância na medicina veterinária e aquicultura. *Biota Amazônia*, 3(1), 109–123. doi:[10.18561/2179-5746/biotaamazonia.v3n1p109-123](https://doi.org/10.18561/2179-5746/biotaamazonia.v3n1p109-123)
- [39] Costa, P. S., Souza, E. Bd, Brito, E. H. Sd, & Fontenelle, R. OdS. (2017). Atividade antimicrobiana e potencial terapêutico do gênero *Lippia sensu lato* (Verbenaceae). *Hoehnea*, 44(2), 158–171. doi:[10.1590/2236-8906-68/2016](https://doi.org/10.1590/2236-8906-68/2016)
- [40] Rodrigues-Soares, J. P., Jesus, G. F. A., Gonçalves, E. L. T., Moraes, K. N., Chagas, E. C., Chaves, F. C. M., ... Martins, M. L. (2018). Induced aerocystitis and hematological parameters in Nile tilapia fed supplemented diet with essential oil of *Lippia alba*. *Brazilian Journal of*

- Veterinary Research and Animal Science, São Paulo*, 55(1), 1–12
- [41] Oliveira, S. R. N., Oliveira, M. A. S., Brandão, F. R., Majolo, C., Chaves, F. C. M., & Chagas, E. C. (2018). Toxicity of *Lippia organoides* essential oil in tambaqui (*Colossoma macropomum*) and its effect against *Aeromonas hydrophila*. *Boletim do Instituto de Pesca*, 44(2), e346.
- [42] Kone, W. M., Solange, K. N. E., & Dosso, M. (2011). Assessing sub-Saharan *Erythrina* forefficity: Traditional uses, biological activities and phytochemistry. *Pakistan Journal of Biological Sciences*, 14(10), 560–571. doi:[10.3923/pjbs.2011.560.571](https://doi.org/10.3923/pjbs.2011.560.571)
- [43] Gratieri-Sossella, A., & Nienow, A. A. (2008). Propagação da corticeira do banhado (*Erythrina crista-galli* L.) (Fabaceae) pelo processo de estaquia. R. *Árvore. Viçosa-MG*, 32(1), 163–171
- [44] Maier, U. H., Rödi, W., Deus-Neumann, B., & Zenk, M. H. (1999). Biosynthesis of *Erythrina* alkaloids in *Erythrina crista-galli*. *Phytochemistry*, 52, 372–382
- [45] Hashimoto, G. (1996). Illustrated Cyclope dia of Brazilian. *Medicinal Plants*. Kamakura, Japan: ABOC-SHA, 678–679.
- [46] Siqueira, M. S., Sousa, R. M., Tirloni, C. A. S., & Gebara, K. S. (2019). CATELAN, T.B.S.; HONORATO, C.A. *Implicações de extratos de Erythrina crista-galli como ansiolítico para Carassius auratus*. *Ciência Animal Brasileira, Goiânia*, 20, 1–8, e-50520.
- [47] Berry, P. E., Hipp, A. L., Wurdack, K. J., Van, E. E. B., & Riina, R. (2005). Molecular phylogenetics of the giants genus *Croton* and tribe *Crotonaeae* (*Euphorbiaceae sensu strictu*) using ITS and trnL-trnF DNA sequence data. *American Journal of Botany*, 92, 1520–1534
- [48] Ramos, J. M. O., Santos, C. A., Santana, D. G., Santos, D. A., Alves, P. B., & Thomazzi, S. M. (2013). Chemical constituents and potencial antiinflammatory activity of the essential oil from the leaves of *croton argyrophyllus*. *Revista Brasileira de Farmacognosia*, 23(4), 644–650. doi:[10.1590/S0102-695X2013005000045](https://doi.org/10.1590/S0102-695X2013005000045)
- [49] Coelho-De-Souza, A. N., Lahlou, S., Barreto, J. E., Yum, M. E., Oliveira, A. C., Oliveira, H. D., ... Leal-Cardoso, J. H. (2013). Essential oil of *Croton zehntneri* and its major constituent anethole display gastroprotective effect by increasing the surface mucous layer. *Fundamental and Clinical Pharmacology*, 27(3), 288–298. doi:[10.1111/j.1472-8206.2011.01021.x](https://doi.org/10.1111/j.1472-8206.2011.01021.x)
- [50] Cavalcanti, J. M., Leal-Cardoso, J. H., Diniz, L. R. L., Portella, V. G., Costa, C. O., Linard, C. F. B., ... Coelho-De-Souza, A. N. (2012). The essential oil *Croton zehntneri* and transanethole improves cutaneous wound healing. *Journal of Ethnopharmacology*, 144, 240–247.
- [51] Souza, E. M., Barbosa, B. S. S., Lorenzo, V. P., Amaral, D. F., Valério, C. S. R. S., Souza, R. C., ... Figueiredo, R. A. C. (2018). REIS. *Uso de extrato bruto de velame Croton heliotropiifolius como aditivo nutricional em juvenis de tilápia do Nilo*. *Revista Semiárido de Visu, Petrolina*, 6(1) (pp. 12–20).
- [52] Francis, J. K. (1989). *Terminaliacatappa*, 4p. Rio Piedras: Institute of Tropical Forestry.
- [53] Costa, C. T. C., Bevilaqua, C. M. L., Morais, S. M., & Vieira, L. S. (2008). Taninos e sua utilização em pequenos ruminantes. *Revista Brasileira de Plantas Mediciniais*, 10(4), 108–116.
- [54] Pinheiro Silva, L., Damacena de Angelis, C., Bonamin, F., Kushima, H., José Mininel, F., ... Akiko Hiruma-Lima, C. (2015). *Terminalia catappa* L.: A medicinal plant from the Caribbean farmacopeia with anti-*helicobacter pylori* and antiulcer action in experimental rodent models. *Journal of Ethnopharmacology*, 159, 285–295. doi:[10.1016/j.jep.2014.11.025](https://doi.org/10.1016/j.jep.2014.11.025)
- [55] Nagappa, A. N., Thakurdesai, P. A., Venkat Rao, N., & Singh, J. (2003). Antidiabetic activity of *Terminalia catappa* Linn fruits. *Journal of Ethnopharmacology*, 88(1), 45–50-50 september. doi:[10.1016/s0378-8741\(03\)00208-3](https://doi.org/10.1016/s0378-8741(03)00208-3)
- [56] Fan, Y. M., Xu, L. Z., Gao, J., Wang, Y., Tang, X. H., Zhao, X. N., & Zhang, Z. X. (2004). Phytochemical and antiinflammatory studies on *Terminalia catappa*. *Fitoterapia*, 75(3–4), 253–260. doi:[10.1016/j.fitote.2003.11.007](https://doi.org/10.1016/j.fitote.2003.11.007)
- [57] Santos, E. L., Souza, A. P. L., Pontes, E. C., Gonzaga, L. S., Ferreira, A. J. S., & de Amendoeira, F. (2015). (*Terminaliacatappa*) como aditivo promotor de crescimento em rações para alevinos de Tilápia do Nilo (*Oreochromis niloticus*). *Agropecuária Técnica*, 36(1), 190–196.
- [58] Claudiano, G. S., Dias Neto, J., Sakabe, R., Cruz, C., Salvador, R., & Pilarski, F. (2009). Eficácia do extrato aquoso de *Terminalia catappa* em juvenis de tambaqui parasitados por monogenéticos e protozoários. *Revista Brasileira de Saúde e Produção Animal*, 10(3) julho/set, 625–636.
- [59] Lorenzi, H., & Brasileira, Á. (2002). *manual de identificação e cultivo de plantas nativas do Brasil*, 2. Odessa: Nova Press: Plantarum.
- [60] Ribeiro, A. B., Silva, D. H. S., & Bolzani, VdS. (2002). Antioxidant flavonol glycosides from *Nectandra grandiflora* (Lauraceae). *Eclética Química*, 27(spe). especial, p. 35–44. doi:[10.1590/S0100-46702002000200004](https://doi.org/10.1590/S0100-46702002000200004)
- [61] Ribeiro, A. B., Bolzani, VdS., Yoshida, M., Santos, L. S., Eberlin, M. N., & Silva, D. H. S. (2005). A new neolignan and antioxidant phenols from *Nectandra grandiflora*. *Journal of the Brazilian Chemical Society*, 16(3B), 526–530. doi:[10.1590/S0103-50532005000400005](https://doi.org/10.1590/S0103-50532005000400005)
- [62] da Silva Filho, A. A., Andrade e Silva, M. L., Carvalho, J. C., & Bastos, J. K. (2004). Evaluation of analgesic and anti-inflammatory activities of *Nectandra megapotamica* (Lauraceae) in mice and rats. *Journal of Pharmacy and Pharmacology*, 56(9), 1179–1184. doi:[10.1211/0022357044058](https://doi.org/10.1211/0022357044058)
- [63] Ferraz, E. O., Vieira, M. A. R., Ferreira, M. I., Fernandes Junior, A., Marques, M. O. M., Minatel, I. O., ... Lima, G. P. P. (2018). Seasonality effects on chemical composition, antibacterial activity and essential oil yield of three species of *Nectandra*. *PLOS ONE*, 13(9), e0204132. doi:[10.1371/journal.pone.0204132](https://doi.org/10.1371/journal.pone.0204132)



- [64] Garlet, Q. I., Rodrigues, P., Barbosa, L. B., Londero, A. L., Mello, C. F., & Heinzmann, B. M. (2019). *Nectandra grandiflora* essential oil and its isolated sesquiterpenoids minimize anxiety-related behaviors in mice through GABAergic mechanisms. *Toxicology and Applied Pharmacology*, 375, 64–80. doi:[10.1016/j.taap.2019.05.003](https://doi.org/10.1016/j.taap.2019.05.003)
- [65] Rodrigues, P., Murari, A. L., Silva, D. T., Peres, M. M., Baldisserotto, B., & Heinzmann, B. M. (2017). Atividade antiparasitária do óleo essencial de *Nectandra grandiflora*: Estudo *in vitro* frente *alchthyophthirus multifiliis*. *Revista Brasileira de Iniciação Científica, Itapetininga*, 4(5).
- [66] Medeiros, E. V. S. S., & Guimarães, E. F. (2007). Piperaceae do Parque Estadual do Ibitipoca, Minas Gerais, Brasil. *Boletim de Botânica*, 25(2), 227–252. doi:[10.11606/issn.2316-9052.v25i2p227-252](https://doi.org/10.11606/issn.2316-9052.v25i2p227-252)
- [67] Duh, C. Y., Wu, Y. C., & Wuang, S. K. (1990). Cytotoxic piperidone alkaloids from the leaves of *Piper aborescens*. *Phytochemistry*, 53, 2689–2691.
- [68] Mamood, S. N. H., Hidayatulfathi, O., Budin, S. B., Ahmad Rohi, G., & Zulfakar, M. H. (2017). The formulation of the essential oil of *Piper aduncum* Linnaeus (Piperales: Piperaceae) increases its efficacy as an insect repellent. *Bulletin of Entomological Research*, 107(1), 49–57. doi:[10.1017/S0007485316000614](https://doi.org/10.1017/S0007485316000614)
- [69] López, K. S. E., Marques, A. M., Moreira, D. L., Velozo, L. S., Sudo, R. T., Zapata-Sudo, G., ... Kaplan, M. A. C. (2016). Local anesthetic activity from extracts, fractions and pure compounds from the roots of *Ottonia anisum* Spreng. (Piperaceae). *Anais da Academia Brasileira de Ciências*, 88(4), 2229–2237. doi:[10.1590/0001-3765201620150821](https://doi.org/10.1590/0001-3765201620150821)
- [70] Cunico, M. M., Trebien, H. A., Galetti, F. C., Miguel, O. G., Miguel, M. D., Auer, C. G., ... de Souza, A. O. (2015). Investigation of local anesthetic and antimycobacterial activity of *Ottonia martiana* Miq. (Piperaceae). *Anais da Academia Brasileira de Ciências*, 87(4), 1991–2000. doi:[10.1590/0001-3765201520140090](https://doi.org/10.1590/0001-3765201520140090)
- [71] Santos, W. B., Majolo, C., Santos, D. S., Rosa, M. C., Monteiro, P. C., Rocha, M. J. S., ... Chagas, E. C. (2018). Eficácia *in vitro* de óleos essenciais de espécies de Piperaceae no controle do acontocéfalo *Neoechinorhynchys buttnerae*. *Revista Brasileira de Higiene e Sanidade Animal*, 12(4), 460–469.